Endothelin receptor B is required for the expansion of melanocyte precursors and malignant melanoma

RONIT LAHAV*
Institut Universitaire de Pathologie, Lausanne, Switzerland

ABSTRACT Since embryonic development and tumorigenesis share common characteristics, studying the role of genes during development can identify molecules that have similar functions in both processes. C-kit and Endothelin receptor B (EDNRB or ETRB) are crucial for melanocyte development in mice and humans but have different functions. While c-kit is needed for survival throughout development until late stages of differentiation in the skin, EDNRB promotes early expansion and migration while delaying the differentiation of melanocyte precursors. Transformation of normal melanocytes to melanoma cells is often associated with gradual loss of differentiation and the gain of high autonomous capacity to proliferate. In accordance with their different roles, c-kit expression is gradually lost during melanoma transformation, while that of EDNRB is greatly enhanced and can serve as a marker of melanoma progression. Inhibiting EDNRB function with a specific antagonist (BQ788) in human melanoma cell lines results in inhibition of growth often associated with induced differentiation indicating that, during melanoma transformation also, the function of EDNRB is to promote growth. EDNRB function does not seem to be essential in the adult, as BQ788 administration to healthy people does not result in major effects. This is probably why BQ788 can specifically inhibit the growth of xenograft human melanoma tumors in nude mice, in a way resembling spontaneous human melanoma regression and why it could serve as a potential therapeutic agent for melanoma.

KEY WORDS: endothelin receptor B, melanocyte, development, melanoma, BQ788

Spots in mice and man

A powerful tool in identifying genes that are important for the development of pigment cells is the characterization of genetic traits that result in white spots due to the lack of melanocytes in the skin. In mice, there are mutations affecting the embryonic development of integumental melanocytes, which typically have no effect on the eyes, but cause a local or global loss of skin pigmentation. These loci affect the cells early in their developmental history and interfere with their ability to migrate and colonize the skin. These strains are called spotting mutants, in which the degree of pigmentation varies from white spots to complete white coat color (and black eye), the latter considered as essentially one big spot (Bennett, 1993). Some of the genes responsible for spotting mutations in mice have a similar function during development of melanocytes in humans since, when mutated, they also result in white areas of the skin in which melanocytes are absent as summarized in Table 1. This review will focus on two ligand receptor systems: c-kit receptor and its ligand Steel or Stem Cell Factor (SCF) and Endothelin receptor B (EDNRB) and its ligand Endothelin 3 (EDN3).

Different spots - different functions

The c-kit – Steel molecular system

KitW or MgfSl heterozygous mice have scattered or diluted spots

In the spotting category, two loci with pleiotropic effects on the development of melanocytes, hematopoietic and germ cell lineages have attracted the interest of embryologists for decades. These are: Dominant White spotting (KitW), identified by Little (Little, 1915) and Steel (MgfSl), first described by Sarvella and Russell (1956) (for reviews see Russell, 1979; Silvers WK, 1979).

By definition, pigmentation is affected in all of the animals heterozygous for any allele at the KitW locus, although the extent of white spotting can vary. The characteristic spotting pattern of KitW heterozygotes is scattered, with no discrete boundaries between pigmented and non-pigmented regions (Geissler et al.,

Abbreviations used in this paper: BQ788, an endothelin receptor B antagonist; EDN3, endothelin 3; EDNRB, endothelin receptor B; MITF, microphthalmia-associated transcription factor; SCF, stem cell factor.

*Address correspondence to: Dr. Ronit Lahav. Institut Universitaire de Pathologie, 25 Rue du Bugnon, Lausanne, Switzerland CH-1011. Fax: +41-21-728-9723. e-mail: ronit.lahav@melcure.com
1981). KitW heterozygotes have a slight dilution of the coat color (Steel received its name from this fact) as well as spotting, white blaze on the forehead, white belt spot and unpigmented feet and tail tip (Servella and Russell, 1956). Viable homozygous of both strains are blacked eye and completely white (Markert and Silvers, 1956; Bennett, 1956). In humans, a high correlation was found between c-kit mutations and piebald patients who suffer from absence of pigmentation from the forehead, eyebrows and chin and of the ventral chest, abdomen and extremities (Fleischman et al., 1991; Spritz et al., 1992; Dippel et al., 1995; Ezoe et al., 1995).

The time at which melanoblasts development diverges from normal in mice carrying KitW or MgfS mutations was not known. Apparently, melanoblasts from KitW homozygotes do not reach the skin since the epidermis is devoid of melanocyte precursors already at 14 days of gestation (Mayer, 1970). Chimeric association of dermal-epidermal tissues from KitW and wild type mice of different embryonic stages led to the suggestion that in KitW the melanocyte precursors are "marginally viable". They are gradually lost during different points in their developmental pathway namely, from the establishment of the line, during the migratory process, colonization of the hair follicles and even after melanin synthesis has occurred (Gordon, 1977; Mayer, 1979).

C-kit is needed during migration, colonization and population of melanoblasts in the skin

The suggestion that KitW melanoblasts are affected at different stages during development until E14 is supported by in situ analysis of the gene product of KitW: c-kit (Geissler et al., 1988; Chabot et al., 1988) and its ligand SCF, the gene encoded by the Steel locus (Martin et al., 1990; Zsebo et al., 1990; Copeland et al., 1990; Williams et al., 1990; Huang et al., 1990). The first few cells expressing c-kit mRNA are detected dorsal to the somites within the migratory pathway of the neural crest cells at E9.5-E10, approximately 24-36 h after crest cells emerge from the neural tube. Expression of c-kit mRNA on presumptive melanoblasts persists thereafter (Manova and Bachvarova, 1991; Yoshida et al., 1996).

In the avian system we could define the exact stage by which neural crest melanocyte precursors start expressing c-kit, by using in situ hybridization in combination with the quail-chick chimera system (Teillet and Le Douarin, 1970; Le Douarin and Kalcheim, 1999). After grafting a quail neural primordium into a chick host, neural crest cells could be followed during the entire migration process. We thus found that neural crest cells migrating along the dorsolateral pathway express c-kit as early as E4, that is about two days after the first neural crest cells have left the neural tube (Lecoin et al., 1995). These results show that c-kit positive cells appear a day or two after neural crest cells start their migration and persist during development.

In mice, using c-kit as a marker for melanoblasts, their progression pattern during different developmental stages could be documented. Analysis of adult KitW heterozygous mice coat color showed that the pattern of melanocyte distribution corresponds to the one observed at E13.5-E14.5 in wild type embryos, meaning that, in KitW mice, melanocytes developed only until that stage (Review: Yoshida et al., 2001).

Functional studies showed that there is a critical requirement for c-kit at E13.5 (Nishikawa et al., 1991) but the lack of c-kit signaling affects melanoblast migration even earlier starting at E11 (Steel et al., 1992)). In addition c-kit was also shown to be required for melanocyte activation in the hair cycle that occurs throughout life (Nishikawa et al., 1991). It seemed, indeed, as was suggested before that c-kit and SCF are needed to sustain melanoblast development in different stages of embryogenesis starting at E10 until E13.5 and also later on during adulthood.

C-kit and SCF are required for melanoblasts survival and differentiation

In order to define the function of SCF, we studied the effect of chicken recombinant SCF on the development of melanocytes from quail neural crest cells in vitro. We showed that SCF mainly promotes survival of neural crest cells and stimulates the differentiation rate of melanocytic precursors (Lahav et al., 1994). These results are in agreement with those showing that SCF is required for survival of murine melanocytes and for their enhanced differentiation in culture (Morrison-Graham and Weston, 1993; Murphy et al., 1992). Taken with the expression pattern, we concluded, similarly to studies carried out in mice, that SCF and c-kit are required mainly for survival and differentiation of melanocyte precursors during migration, colonization and population of the skin.

The Endothelin System

In EdnrbS-l and Edn3ls mice, spotted areas have defined margins

Piebald is a recessive mutation known in the mouse since a long time. Homozygotes have dark eyes and show irregular white spotting of the coat, especially on the belly, sides and back. The borders between white and pigmented areas are sharply defined. The white areas of the coat completely lack melanocytes. There is a reduction in the number of melanocytes in the choroid layer of the eye. In addition, homzygotes may develop megacolon that is always associated with lack of ganglion cells in the distal portion of the colon. Mice with a more severe mutation at the piebald locus, piebald lethal (EdnrbS-l), have dark eyes and an almost completely white coat with pigmented hair restricted to small areas on the head and base of the tail. All EdnrbS-l homozygotes develop megacolon with lack of enteric ganglion cells in the posterior end of the colon. They usually die at about two weeks of age, but some live a year or more and may breed (Lyon and Searle, 1990).

Another recessive mutation that resembles piebald mice is Lethal spotting (Edn3ls). Lethal spotting mice usually die in the third...
EDNRB in melanocyte development and cancer

EDNRB is needed before the onset of migration

Murine and avian neural crest cells start expressing EDNRB at the premigratory stage (Reid et al., 1996; Nataf et al., 1996). Avian EDNRB is detected from E2 when crest cells become individualized in the dorsal neural primordium, which is about two days before the onset of c-kit expression. Avians have a second receptor, EDNRB2, which later on is only expressed by melanoblast and melanocytes (Lecoin et al., 1998). Both receptors seem to be important for melanocyte development since stimulation of cultured neural crest cells with EDN3 results in an initial increase in the expression of EDNRB that is followed by enhanced expression EDNRB2 (Lahav et al., 1998).

EDNRB also affects the melanogenic lineage at earlier developmental stages than c-kit. Murine melanoblasts were shown to be affected in Ednrbs-1 mutants before they express melanogenic markers (Pavan and Tilghman, 1994) and not after as was shown for c-kit mutants. In addition analysis of the coat pigmentation pattern of EDNRB deficient mice shows that it resembles the distribution pattern of E10 embryos (Yoshida et al., 2001), meaning that, in these mice, melanocyte precursors progressed in their development only until this stage and not until E13.5 as in c-kit deficient mice. The period between E10 and E12.5 was shown to be critical for melanoblasts development since stimulation of cultured neural crest cultures strikingly increased the number of melanocytes. Examining the evolution of such cultures revealed that this was a result of a massive increase in proliferation accompanied by an initial delay of differentiation. Repeating these experiments in defined medium conditions showed clearly that the functions that could be attributed to EDN3 alone are indeed enhancement of proliferation and inhibition of differentiation. These results are in agreement with those of Sviderskaya et al. (1998) showing that primary melanoblast cultures taken from EDNRB mutant mice were differentiation-deficient compared with wild-type.

We could also show that EDN3 affects neural crest precursors before they are committed to the melanocytic lineage. Clonal cultures showed that EDN3 expands the common precursors of melanocytes and glia cells (Lahav et al., 1998). Moreover addition of EDN3 to melanocyte or glia cultures can revert the cells to the earlier common precursor stage resulting in the appearance of both melanocytes and glia from clonal population of either melanocytes or glia (Dupin et al., 2000; Dupin and Le Douarin, 2003; Dupin et al., 2003).

The big spot picture

Some melanoblasts die before they migrate, others die after

Taken together it seems that the EDNRB/EDN3 receptor-system is needed to promote early precursor proliferation before or coincident with neural crest migration while the c-kit/SCF system is needed for survival and differentiation of melanocyte precursors while they colonize the embryonic skin.

The functional difference between these two molecular systems is also reflected in the different spotting patterns between Ednrbs-1 and KitW- mice.

To explain spotting, Mintz proposed that the whole coat pigmentation derives from 34 melanoblasts according to the maximum number of territories that she could observe in aggregation chimeras (Mintz, 1967). Each melanoblast migrates and proliferates along the dorso-ventral axis to populate a strip-like territory. Spots

Week of life, however some survive and are fertile. Homozygotes have considerable white spotting and megacolon. Like in piebald, the most distal portion of the bowel is aganglionic in these mice.

The breakthrough in the study of the developmental abnormalities caused by Ednrbs-1 and Edn3- mice has been the identification of the genetic defects responsible for these mutations and the discovery that a related defect is also present in a subset of patients with Hirschsprung’s disease (Paffenberger et al., 1994; Hosoda et al., 1994; Baynash et al., 1994) for review see (McCallion and Chakravarti, 2001). These abnormalities are in genes encoding EDNRB or its ligand EDN3.

The endothelins 1, 2 and 3 (EDN1, EDN2 and EDN3) are a family of 21 amino acid peptides that activate one or both of the two heptahelical, G-protein-coupled endothelin receptors, A and B (EDNRA and EDNRB). EDNRA exhibits different affinities for endothelin peptides with a potency rank order of EDN1>EDN2>EDN3 (Yanagisawa, 1994). EDNRB accepts all three peptides equally (Sakurai et al., 1990), as shown in Fig. 1.
occur when the descendants of certain clones die since their progenitors were «preprogrammed» to die. Since their death happens relatively late, other clones cannot expand to their territory and an unpigmented region occurs.

Schaible suggested that there are 14 melanoblasts, each migrating to a center of a territory and expanding. Spotting occurs when the early melanoblasts die or cannot proliferate (Schaible, 1969). Other clones cannot expand to cover the depigmented area since this process is limited in time. Clones can expand before the tissue environment differentiates to the point that it restricts pigment cell migration.

Our data suggest that between the stages at which melanoblasts require EDNRB and c-kit function, a large expansion in population size occurs. This is supported by experiments showing that murine EDNRB function is required before E10.5. At that stage, very few melanoblasts are detected in wild type mice as suggested by Mintz and Schaible (Pavan and Tilghman, 1994). By the time that c-kit is first needed (E11.5) the dermis is seeded with numerous cells identified as melanoblasts (Wehrle-Haller and Weston, 1995).

Taken together, spotting in \( Ednrb^{Sl} \) and \( Edn3^{Sl} \) might occur according to the explanation provided by Schaible since the precursors are probably affected before they expand. In contrast in \( Kit^{Sl} \) or \( Mgf^{Sl} \), the mechanisms that lead to spotting resemble more those described by Mintz where melanoblasts that are destined to die (i.e. carry a mutation) first expand. These differences might also account for the differences in the nature of spots. In \( Ednrb^{Sl} \) and \( Edn3^{Sl} \), the spots have sharp margins since the precursors were eliminated early resulting in the absence of all their descendants in a certain area of the skin (Fig. 2 right). In contrast, in \( Kit^{Sl} \) or \( Mgf^{Sl} \), melanoblasts are affected relatively late. At this stage many melanoblasts are found in the dermis. Partial elimination of melanoblasts would result in small distances among pigmented and unpigmented hair (Fig. 2 left). This pattern is well represented in mice that were treated relatively late (E13.5-E14.5) with an antibody which antagonizes c-kit function (Nishikawa et al., 1991).

Interestingly melanoblasts show an uneven distribution along the murine neural axis. Many more melanocyte precursors are found in the head and tail as compared to the trunk region (until E15.5) consistent with the areas that are often pigmented in spotted mice (Pavan and Tilghman, 1994; Wehrle-Haller and Weston, 1995). This could explain why pigmentation is more often retained in the head and tail regions.

**Spot signaling:** C-kit can promote survival and differentiation by activating MITF. EDNRB interactions are not yet known

One of the molecular pathways by which c-kit and SCF affect melanocyte development involves genes with known effects on pigmentation in mice and man, like: MITF, SOX10 and Pax3 (see table 1) (Review: Rawls et al., 2001). C-kit can affect differentiation and survival of melanocyte precursors by inducing MITF phosphorylation and transcriptional activity (fig. 3). MITF (microphthalmia-associated transcription factor) is a basic helix-loop-helix/leucine zipper protein that is considered as a melanocytic key regulator since its overexpression can cause cells to adopt melanocyte characteristics (Tachibana et al., 1996; Lister et al., 1999). MITF has been shown to induce the transcription of pigmentation genes like tyrosinase and survival factors like BCL2 (Hou et al., 2000; McGill et al., 2002). In this model, a downstream component of the kit-signaling pathway, MAPK, phosphorylates MITF, thereby creating a binding site for the transcriptional coactivator p300 (Sato et al., 1997; Hemesath et al., 1998; Price et al., 1998). This transcriptional activation will stimulate MITF before its rapid ubiquitin-mediated degradation (Wu et al., 2000).

EDNRB interactions with MITF, Pax3 and SOX10 are not well understood in spite of the fact that mutations in EDNRB and SOX10 cause exactly the same syndrome in mice and SOX10 cause exactly the same syndrome in humans and mice (Southard-Smith et al., 1998; Potter et al., 2000). Recently, SOX10 was shown to have multiple binding sites on an EDNRB enhancer which serve to up-regulate EDNRB expression in the enteric system (Zhu et al., 2004). In addition, EDNRB inhibition causes down regulation of its own expression in melanomas, suggesting that EDNRB signaling can enhance its own expression (Lahav et al., 2004). Taken together, since EDNRB is needed in order to expand melanocyte-glia precursors while delaying their differentiation, one could suggest the following sequence of events: the EDNRB signaling pathway functions first, amplifying itself with the continuous presence of its ligand EDN3 and the transcription factor SOX10. Once enough precursors are produced, MITF

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**Fig. 3.** c-kit can promote survival and differentiation by activating the transcriptional activity of MITF which will induce the expression of BCL2 and Tyrosinase. One way by which MITF transcription can be regulated is by SOX10 and Pax3. EDNRB interaction in this pathway is suggested by the fact that mutations in both EDNRB and SOX10 cause the same syndrome in humans, although the mechanism is unknown.
transcription is transiently stabilized by c-kit to allow the gradual differentiation and enhanced survival of melanocyte precursors.

**Spots and melanoma**

**From no melanocytes to too many**

The temporal and functional differences between the c-kit/SCF and EDNRB/EDN3 receptor-ligand systems in their effect on melanocyte precursors seems to be reflected also in the situation of melanocyte malignancy. C-kit expression is normally reduced or absent in melanoma cell lines and it seems therefore, that a reduction in c-kit gene expression either promotes or is a consequence of transformation of melanocytes (Lassam and Bickford, 1992; Zakut *et al.*, 1993) for review (Nyormoi and Bar-Eli, 2003). This finding correlates well with loss of differentiation function characterizing the malignant phenotype. In contrast melanoma cells acquire a high capacity to proliferate (Review: Halaban *et al.*, 2000) since EDNRB promotes proliferation while inhibiting differentiation, it is not surprising that its expression is highly enhanced in cutaneous melanoma (Loftus *et al.*, 1999). In a study that examined gene expression profiling of 6,971 genes in 31 human melanoma samples from biopsies or tumor cell cultures, a general increase of EDNRB expression was observed (Bittner *et al.*, 2000). A similar observation was made in another study of gene expression patterns in human cancer cell lines. Here again, all the melanoma samples tested showed overexpression of EDNRB (Ross *et al.*, 1999). EDNRB was also found to be a melanoma progression marker since immunohistochemistry on paraffin-embedded tissue sections of 159 human melanoma cases revealed an increase of EDNRB expression as melanoma progressed to metastatic disease (Demunter *et al.*, 2001).

These findings suggest that EDNRB activation contributes to melanoma development and progression. In support of this view is the finding that EDN1 is secreted by keratinocytes in the skin in response to ultraviolet irradiation (Imokawa *et al.*, 1992; Ahn *et al.*, 1998), which is known to be the most common causative factor of the disease (Atillasoy *et al.*, 1998). Moreover, UV-induction of keratinocyte EDN1 down regulates E-cadherin in melanocytes and melanoma (Jamal and Schneider, 2002). Down regulation of E-cadherin expression is typical for melanomas and serves to enhance their invasive capability (Hsu *et al.*, 2000).

**Treating melanoma**

In order to assess the functional importance of EDNRB for melanoma progression, we used a specific antagonist of the receptor: BQ788 (Ishikawa *et al.*, 1994). BQ788 is a highly selective inhibitor of EDNRB that was developed as a potential drug for cardiovascular disorders since endothelins serve in the adult as potent vasoconstrictors. However, these effects are predominantly mediated by EDNRA (for review: Luscher and Fig. 4. Effect of the EDNRB antagonist BQ788 on neural crest cultures treated with EDN3. Neural crest cultures were treated with EDN3 for 5 days during which there is a massive increase in cell number and inhibition of differentiation (A). Administration of BQ788 results in decrease in cell number and induced differentiation as can be seen by the appearance of pigmented cells (B).

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**TABLE 1**

**SPOTTING IN MAN AND MICE**

<table>
<thead>
<tr>
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<th>Symptoms</th>
<th>Mouse mutant</th>
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<td>Ednrbs-l&lt;sup&gt;+/−&lt;/sup&gt;: Piebald (spotting)</td>
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<td>Pigmentary disturbance and high frequency of cochlear deafness.</td>
<td>Mitf&lt;sup&gt;−/−&lt;/sup&gt;: Whit coat, abnormal eye, osteoporosis, hematopoietic and neural crest defects.</td>
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Viability and enhance differentiation when treated with BQ788. In contrast, treatment of kidney 293 cells dramatically and treatment of SK-MEL-5 cells also results in an increase in pigmentation (Fig. 5A,C). Cell numbers reduce et al., healthy volunteers left their blood pressure unaffected (Strachan Barton, 2000)). This is probably why administration of BQ788 to human kidney cells. Fig. 5. Effects of the EDNRB antagonist BQ788 on human melanoma cells. Human melanoma cells SK-MEL-28 (B) and SK-MEL-5 (D) loose viability and enhance differentiation when treated with BQ788 (A,C). Human kidney cells (F) do not seem to be affected by BO788 (E).

To test BQ788 effectiveness in inhibiting tumor growth in vivo, human melanoma cell lines were induced to grow as subcutaneous tumors in nude mice. About two weeks later when the tumors were growing stably, the mice were divided into two groups receiving either vehicle or BQ788 injected daily into the tumor. During 10 days of treatment a significant and reproducible inhibition of tumor growth is observed in the BQ788 treated groups of mice. Systemic administration of the drug under the same conditions results in a significant reduction in tumor growth, with however a greater variability in the response of the treated group. Further analyses show that the treated mice can be divided into two groups, half with significant but moderate inhibition of tumor growth and the others with an almost complete shrinkage of the tumors. One of the reasons for the inhibition of tumor growth is enhanced apoptosis as revealed by a general increase in tunnel staining in tumor sections (Lahav et al., 1999).

Recently, our observation has been corroborated with a different EDNRB antagonist showing that EDNRB inhibition is effective in inhibiting human melanoma tumor growth in nude mice (Bagnato et al., 2004). Moreover, using BQ788, the authors could show that EDNRB mediates many known molecular events characteristic of melanoma progression (Bagnato et al., 2004).

As melanoma is of more metastatic grade, it shows higher EDNRB expression and as a result is more sensitive to EDNRB inhibition resulting in increased apoptosis. This suggests that EDNRB inhibition would be most effective against metastatic melanoma. EDNRB expression levels are also important for melanoma survival. Specific reduction in RNA and protein levels of EDNRB using RNAi leads to reduced melanoma viability (Lahav et al., 2004).

Studying the mechanisms by which EDNRB induces apoptosis in melanoma has revealed the participation of two genes, BCL-2A1 and PARP-3, with known functions in survival and apoptosis. We show that EDNRB inhibition results in down regulation of BCL-2A1 and PARP-3 before the cells start dying, and levels of reduction in their expression directly correlate with the levels of apoptosis induced (Lahav et al., 2004). Therefore it seems that advanced, metastatic melanoma expresses higher levels of EDNRB and inhibition of EDNRB function leads to reduced expression levels of EDNRB, BCL-2A1 and PARP-3 and subsequently to apoptosis.

Interestingly, EDNRB inhibition up-regulates VEGF and other angiogenic factors resulting in increased appearance of blood vessels in BQ788 treated tumors with inhibited growth (Lahav et al., 2004). This is a feature which fits well with the pathology of spontaneously regressing human melanomas that are characterized by increased vascularity (Weedon, 2002). Thus, EDNRB inhibition may also be the driving force of human melanoma regression highlighting even more the therapeutic potential of EDNRB inhibition for melanoma.

References


EDNRB in melanocyte development and cancer


