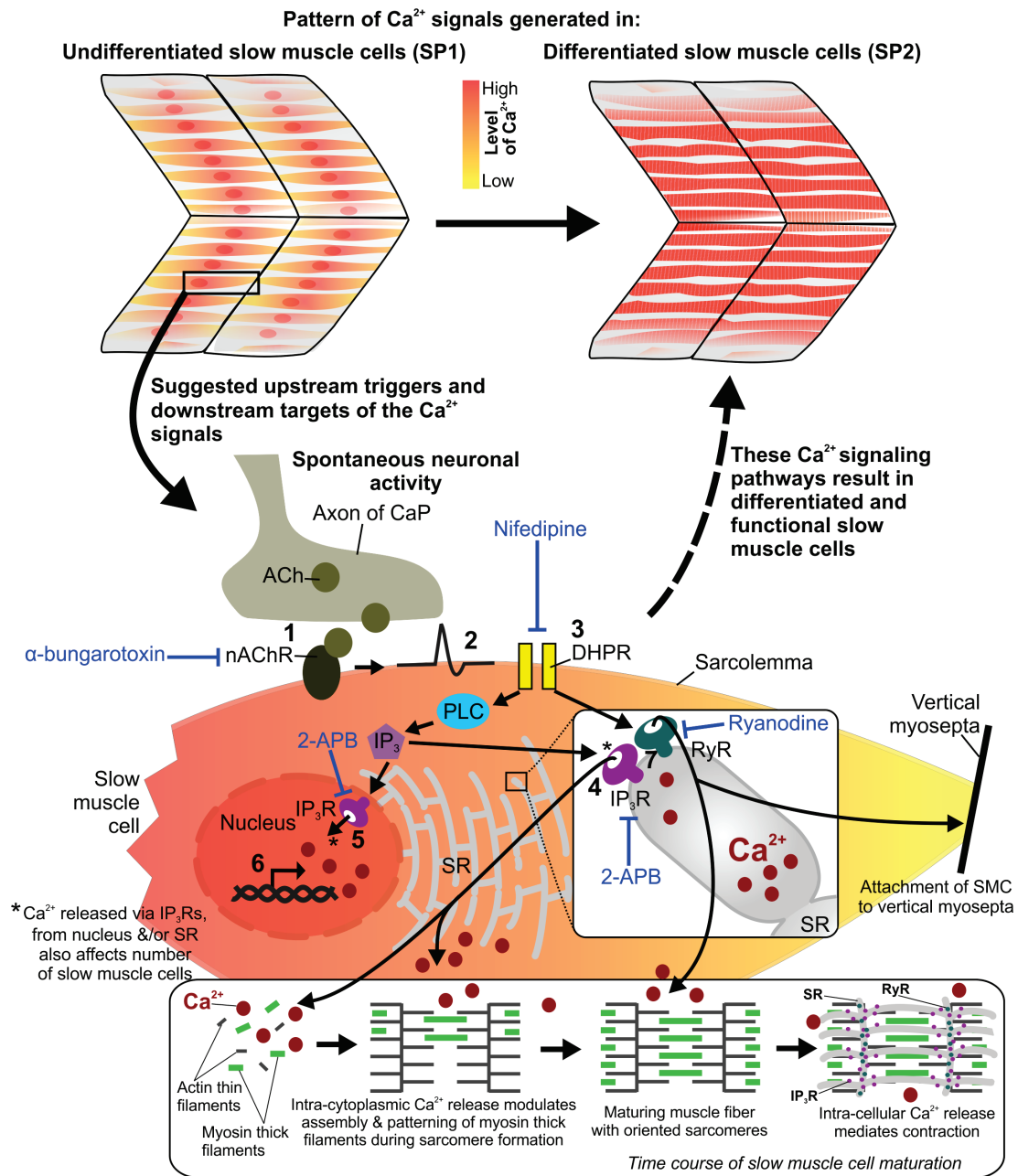


SUPPLEMENTARY MATERIAL

corresponding to:

**Visualization, characterization and modulation of
calcium signaling during the development of
slow muscle cells in intact zebrafish embryos**

CHRIS Y. CHEUNG, SARAH E. WEBB, DONALD R. LOVE and ANDREW L. MILLER*



Supplementary Fig. S1. Schematic diagram of the proposed signaling cascade for the generation of the SP1 Ca²⁺ signals in SMCs during myofibrillogenesis. At ~17.5 hpf, (i.e., when the SP1 Ca²⁺ transients begin), the myotomes are innervated by the caudal primary (CaP) axons (Eisen et al., 1986; Myers et al., 1986; Westerfield et al., 1986). Acetylcholine (ACh) released from the axon binds to nicotinic acetylcholine receptors (nAChRs) on the surface of the SMCs (1), and thus induces a depolarization of the sarcolemma (2). This depolarization activates dihydropyridine receptors (DHPRs) (3), that in turn stimulate the release of Ca²⁺ (see red circles) from intracellular stores, i.e., the sarcoplasmic reticulum (SR) and perinuclear cisternae. The SP1 Ca²⁺ signals are generated by Ca²⁺ release via both IP₃Rs and RyRs. It has been proposed that DHPRs can activate the production of IP₃ from phospholipase C (PLC) and then induce Ca²⁺ release via IP₃Rs from the SR (4) and the perinuclear cisternae (5); (Jaimovich et al., 2000; Powell et al., 2001; Araya et al., 2003). Moreover, it has been shown that elevations of nuclear Ca²⁺ can activate gene expression in skeletal muscle cells (6) (Carrasco et al., 2003; Cárdenas et al., 2005). We suggest that the Ca²⁺ released via IP₃Rs might play a role in regulating the expression of some important proteins that are involved in sarcomere assembly. In addition, it has been proposed that direct coupling of DHPRs with juxtaposed RyRs can trigger Ca²⁺ released through RyRs (7); (Schneider and Chandler, 1973). We suggest that the Ca²⁺ released via RyRs maintains the integrity of the sarcomeres and regulates muscle contraction. Inhibiting the nAChRs, DHPRs, IP₃Rs and RyRs with α -bungarotoxin, nifedipine, 2-APB and ryanodine, respectively (indicated in blue) can block the SP1 signals generated in SMCs and thus disrupt different aspects of muscle development.

Movie 1 Representative example of the Ca²⁺ signals generated in some of the slow muscle cells in somite 8 (S8) of a wild-type embryo at ~17.5 hpf (i.e., the 17-somite stage), as visualized by confocal microscopy using calcium green-1 dextran. Anterior is to the left and dorsal is to the top.