doi: 10.1387/ijdb.082793jr



## SUPPLEMENTARY MATERIAL

corresponding to:

## Eye-specific expression of an ancestral jellyfish *PaxB* gene interferes with *Pax6* function despite its conserved Pax6/Pax2 characteristics

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Supplementary Fig. 1. Normal gross morphology of the control LR-Cre transgenic eyes. Schematic diagram of LR-Cre (A) and PaxB (B) transgenic constructs. Three copies of the lens-specific ectoderm enhancer (EE) derived from the Pax6 gene were cloned upstream of the Pax6 P0 minimal promoter to drive the expression of Cre and EGFP (A; LR-Cre Tg) or the expression of PaxB and Cre cDNAs (B; PaxB Tg) in bicistronic constructs via internal ribosomal entry site (IRES). Cre recombinase activity in LR-Cre (C) and PaxB transgenic (D) mice (detected using ROSA26R reporter mice) at E10.5. In comparison to PaxB transgenic mice the Cre recombinase activity in LR-Cre transgenic mice is much stronger in all developing eye tissues. Note the intensive staining in the presumptive retina of LR-Cre transgenic mice. Cryosections at the indicated embryonic stages from the LR-Cre (E,G) and PaxB transgenic (F,H) mice stained with hematoxylin and eosin. The morfology of E11.5 and E15.5 LR-Cre transgenic eyes appears normal as compared to PaxB transgenic mice despite stronger expression of Cre. Abbreviations used in this are as follows: I, lens; Ip, lens pit; pr, presumptive retina.



Supplementary Fig. 2. Lens specific transcription factors and crystallins are reduced in *PaxB* transgenic mice. Cryosections of wild-type (A,B,C,D,E,F,G) and PaxB transgenic (A',B',C',D',E',F',G') embryos at E11.5 stained for Prox1 (A,A'), cMaf(B,B'),  $\beta$ -crystallin (C,C'),  $\alpha$ -crystallin (D,D'),  $\beta$ A3-crystallin (E,E'),  $\gamma$ -crystallin (F,F') and MIP26 (G,G'), and counterstained with DAPI (blue). Lower immunoreactivity of all above mentioned markers and smaller lens was observed in the PaxB transgenic mice.



Supplementary Fig. 3. Transcription factors important for the eye development were expressed normally in *PaxB* transgenic mice. Cryosections of wild-type (A,C,E,G,I,K) and PaxB transgenic (B,D,F,H,J,L) embryos at E10.5 were stained for Pax6 (Covance, A,B), Pax2 (C,D), Vsx2 (Chx10) (E,F), Six3 (G,H), Meis1 (I,J) and Meis2 (K,L), counterstained with DAPI (blue). Pax6, Pax2 and Vsx2 (Chx10) staining appeared normal in the PaxB transgenic mice. No difference was observed in the expression Six3, Meis1 and Meis2, transcription factors acting upstream of Pax6.



Supplementary Fig. 4. The photoreceptor cells were not affected in the *PaxB* transgenic retina. Immunostaining of the PaxB transgenic (right panels) and wild-type (left panels) adult retinal sections. Applied antibodies were as follows: CRX (cone-rod homeobox containing gene, A-B'), NR2e3 (protoreceptor specific nuclear receptor, C-D'), M-opsin (M-cone photoreceptors, E-F'), S-opsin (S-cone photoreceptors, G-H'), arrestin (rod-photoreceptors, I-J'), rhodopsin (rod-photoreceptors, K-L'), GRK1 (rhodopsin kinase, M-N'). The nuclei were counterstained with DAPI. No difference was apparent in the photoreceptor cells using a variety of photoreceptor markers between wild-type and PaxB transgenic retinas. Abbreviations used in this figure are as follows: gcl, ganglion cell layer; inl, inner nuclear layer; ipl, inner plexiform layer; onl, outer nuclear layer; opl, outer plexiform layer; rpe, retinal pigment epithelium.